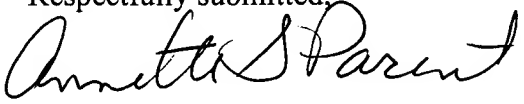
If the Examiner believes a telephone conference would expedite prosecution of this application, please telephone the undersigned at 415-576-0200.

Respectfully submitted,



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VERSION WITH MARKINGS TO SHOW CHANGES MADE

In the Specification:

Paragraph beginning at line 26 of page 29 has been amended as follows:

As a first step, experiments were carried out to determine what are the optimum conditions for immobilizing a capture oligonucleotide to a plate. In this experiment, Capture Oligonucleotide1 (5'ACCGCACCCGCTCCGTCCCATTGAAGAAAT; SEQ ID NO:278) was modified with an amine at the 5' end with a C6 linker and a biotin group on the 3' end. For the purpose of actual HLA genotyping, the Capture Oligonucleotide will not have a biotinylated 3' end. The oligonucleotide1 was incubated on a 96 well Covalent Binding Microwell plate (Xenobind™, Xenopore, Hawthorne, NJ) according to the manufacturer's instructions. The plate was then washed three times with phosphate-buffered-saline (PBS). ExtrAvidin® Peroxidase (SIGMA) was added and allowed to incubate on the tray. The plate was washed three times with PBS. TMB substrate (3,3',5,5' – Tetramethylbenzidine) was added to the plate, 1N HCl added and tray was read at 450 nm. The current optimum conditions for oligonucleotide binding was Capture Oligonucleotide at 100 ng/ul in PBS at pH 8.8 incubated overnight at 4°C. Alternatively, binding can occur at 37 °C for 2 hours with Capture Oligonucleotide at 100 ng/ul in PBS at pH 8.8.